

ORIGINAL ARTICLE

HERBase: A collection of understory herb vegetation plots from Amazonia

Thiago ANDRÉ¹, Gabriel Massaine MOULATLET^{2*}, Thaís Elias ALMEIDA³, Paula Palhares de Polari ALVERGA⁴, Carlos Renato BOELTER⁵, Debora Pignatari DRUCKER⁶, Julia Gomes da SILVA⁴, Reynaldo LINARES-PALOMINO⁷, Maria Aparecida LOPES⁸, José Leonardo Lima MAGALHÃES⁹, Angelo Gilberto MANZATTO¹⁰, Henrique Augusto MEWS¹¹, Iracema Elizabeth de Siuza MOLL¹², Amanda Frederico MORTATI¹³, Eliana Celestino da PAIXÃO¹⁴, Estela QUINTERO-VALLEJO¹⁵, Tinde van ANDEL¹⁶, Marcos SILVEIRA⁴, Danielle STORCK-TONON¹⁷, Hanna TUOMISTO¹⁸, Gabriela ZUQUIM^{18,19}, Flávia Regina Cappelloto COSTA⁵

¹ Universidade de Brasília, Instituto de Ciências Biológicas, Departamento de Botânica, Brasília, Distrito Federal, Brazil

² Instituto de Ecología A.C., Red de Biología Evolutiva, Xalapa, Veracruz, Mexico

³ Universidade Federal de Pernambuco, Centro de Biociências, Departamento de Botânica, Recife, Pernambuco, Brazil

⁴ Universidade Federal do Acre, Rio Branco, Acre, Brazil

⁵ Instituto Nacional de Pesquisas da Amazônia (INPA), Manaus, Amazonas, Brazil

⁶ Embrapa Agricultura Digital, Campinas, São Paulo, Brazil

⁷ Smithsonian's National Zoo and Conservation Biology Institute, Washington, DC, USA

⁸ Universidade Federal do Pará, Belém, Pará, Brazil

⁹ Universidade Federal do Amapá, Macapá, Amapá, Brazil

¹⁰ Universidade Federal de Rondônia, Porto Velho, Rondônia, Brazil

¹¹ Universidade Federal de Rondonópolis, Instituto de Ciências Exatas e Naturais, Rondonópolis, Mato Grosso, Brazil

¹² Secretaria de Meio Ambiente e de Políticas Indígenas (SEMAPI), Rio Branco, Acre, Brazil

¹³ Universidade Federal do Oeste do Pará, Instituto de Biodiversidade e Florestas, Santarém, Pará, Brazil

¹⁴ Instituto Nacional de Ciência e Tecnologia em Áreas Úmidas - INAU, Cuiabá, Mato Grosso, Brazil

¹⁵ Universidad CES, Medellín, Antioquia, Colombia

¹⁶ Naturalis Biodiversity Center, Leiden, the Netherlands

¹⁷ Universidade do Estado de Mato Grosso, Programa de Pós-graduação em Ambiente e Sistemas de Produção Agrícola, Tangará da Serra, Mato Grosso, Brazil

¹⁸ University of Turku, Department of Biology, Turku, Finland

¹⁹ Aarhus University, Section for Ecoinformatics and Biodiversity, Department of Biology, Aarhus, Denmark

* Corresponding author: gabriel.moulatlet@gmail.com; <https://orcid.org/0000-0003-4148-3662>

ABSTRACT

Understorey herbs form a diverse and understudied plant assemblage in tropical forests. Although several studies and research teams have long been dedicated to the study of this conspicuous vegetation component in Amazonia, no effort to unify the data has been undertaken to date. In contrast to trees and other life forms for which major data compilations already exist, a unified database dedicated to herbs is still lacking. Part of the problem is in defining what is a herb and how to effectively sample herb assemblages. In this article, we describe the database HERBase, an exhaustive compilation of published and unpublished data on herb inventories in Amazonia. We also describe the structure, functioning, and guidelines for data curation and integration in HERBase. We were able to compile information from 1381 plots from all six Amazonian geographic regions. Based on this dataset, we describe and discuss sampling and knowledge gaps, priority areas for new collections, and recommend sampling protocols to facilitate data integration in the future. This novel database provides a unique biodiversity data repository on understorey herbs that will enable new studies on community ecology and biogeography.

KEYWORDS: angiosperms, database, ferns, tropical forests, Zingiberales

HERBase: Uma coleção de parcelas de ervas de sub-bosque da Amazônia

RESUMO

As ervas do sub-bosque formam um componente diversificado e pouco estudado em florestas tropicais. Embora vários estudos e grupos de pesquisa tenham se dedicado ao estudo desse componente conspícuo na Amazônia, nenhum esforço foi feito até o momento para unificar essas informações. Em contraste com árvores e outros grupos de plantas para os quais já existem grandes compilações de dados, uma base de dados unificada dedicada às ervas ainda não existe. Parte do problema está em definir o que é uma erva e como amostrar comunidades de ervas de forma eficiente. Neste artigo descrevemos a base de dados HERBase, uma compilação exaustiva de dados publicados e não publicados sobre inventários de ervas na Amazônia. Também descrevemos a estrutura, funcionamento e diretrizes para curadoria e integração de dados na HERBase. Conseguimos compilar informações de 1381 parcelas de todas as seis regiões geográficas amazônicas. Com base nesses dados, descrevemos e discutimos lacunas de amostragem e conhecimento, apontamos áreas prioritárias para novas coletas e recomendamos protocolos de amostragem para facilitar a integração de dados no futuro. Essa nova base de dados fornece dados únicos de biodiversidade sobre ervas do sub-bosque que permitirão novos estudos sobre ecologia e biogeografia de comunidades.

PALAVRAS-CHAVE: angiospermas, base de dados, samambaias, florestas tropicais, Zingiberales

CITE AS: André, T.; Moulatlet, G.M.; Almeida, T.E.; Alverga, P.P.P.; Boelter, C.R.; Drucker, D.P.; Silva, J.G.; Linares-Palomino, R.; Lopes, M.A.; Magalhães, J.L.L.; Manzatto, A.G. *et al.* 2023. HERBase: A collection of understory herb vegetation plots from Amazonia. *Acta Amazonica* 53: 114-121.

INTRODUCTION

The study of herb ecology and distribution in tropical forests has advanced in recent decades (e.g., Cicuzza *et al.* 2013; Tuomisto *et al.* 2003a, 2019; Figueiredo *et al.* 2022) but is still lagging behind studies of other biological components (Perea *et al.* 2022), such as trees (ter Steege *et al.* 2003, 2006, 2013; Draper *et al.* 2021) and birds (Ribas *et al.* 2005). It is still an open question to what degree ecological theories and generalizations derived from trees apply to tropical forest herbs (e.g., Janzen-Connell mechanisms). Although canopy trees define forest structure and control most of the carbon and water fluxes, understory plants such as herbs and shrubs are an important part of the forest ecosystem and contain a large proportion of its taxonomic diversity (Dodson and Gentry 1978, Ribeiro *et al.* 1999). For instance, herbs were found to contribute about 30% of the total species richness in the Rio Palenque Reserve in Ecuador (Dodson and Gentry 1978), and 23% in the Ducke Reserve in Brazil (Ribeiro *et al.* 1999). This taxonomic diversity promotes varied and complex interactions with animals (Royo and Carson 2006), and can also be expected with microorganisms (which are largely unknown). Herbs can also affect forest regeneration, as they compete with tree seedlings for space and resources and can thereby filter tree species composition or even prevent or delay tree establishment (George and Bazzaz 1999). However, all these relationships are little studied.

Much of the ecological knowledge using floristic inventories on tropical forest plants has been obtained from studies of trees (Hubbell 2001; ter Steege *et al.* 2013). Trees are laborious to sample. Instead, general floristic patterns could be studied by focusing on understory plants. Species turnover patterns of at least ferns and lycophytes have been found to be rather similar to those of trees (Ruokolainen *et al.* 1997, 2007; Vormisto *et al.* 2000; Jones *et al.* 2008, 2013; Higgins *et al.* 2011; Tuomisto *et al.* 2016). Knowledge of community composition, as well as of the distribution and niches of species, is fundamental for the construction of macroecological and biogeographic hypotheses, and to support conservation programs (Hortal *et al.* 2015). Inadequate and biased sampling has adverse effects on our understanding of biodiversity patterns (Moerman and Estabrook 2006; Grand *et al.* 2007; Yang *et al.* 2013; Oliveira *et al.* 2016a). Recognized shortfalls of biological knowledge include the Linnaean shortfall (many species remain taxonomically undescribed and unnamed; Whittaker *et al.* 2005), the Wallacean shortfall (lack of knowledge on the geographic distribution of species; Whittaker *et al.* 2005), and the Hutchinsonian shortfall (lack of knowledge on the ecological niches of species; Hortal *et al.* 2015). Field studies documenting biological diversity have been carried out in a highly biased way, such that some regions of the globe and some biological lineages have been intensively

studied while little information exists for others (Whittaker *et al.* 2005; Diniz-Filho *et al.* 2010; Cornwell *et al.* 2019).

Inventories based on plot data can greatly reduce such shortfalls of biological knowledge, especially when implemented through standardized and integrated sampling (Magnusson 2013). Combining efforts to build large databases can boost biodiversity research, reduce the aforementioned shortfalls and enable the macroscale analyses required to understand a world under global change (Magnusson 2019). Much of the progress in the understanding of tree, palm, and liana ecology in the last two decades has been due to the creation of collaborative networks, which have either applied standardized protocols to collect biodiversity and demographic data [e.g., CTFS (<http://ctfs.si.edu/>); PPBio (<https://ppbio.inpa.gov.br/>); RAINFOR (<https://rainfor.org/>)] or have compiled existing but dispersed data into accessible repositories [e.g., ATDN (<https://www.atdn.myspecies.info/>); DRYFOR (<http://www.dryflor.info/>); NeoTropTree (<http://www.neotropree.info/>)]. Standardised data on ferns and lycophytes across Amazonia have been compiled by the Amazon Research team of the University of Turku (Finland) (www.utu.fi/amazon), but data on the distribution and abundance of other tropical forest herbs have not yet been organised. To fill this gap, we started the Research Network on Amazonian Understorey Herb Communities (HERBase). Its purpose is to assemble data on the abundances of herb species in sampling plots that have a known size and, if available, also associated environmental data. HERBase provides the opportunity to address broad-scale questions, fostering the understanding of herb ecology, evolution, systematics, conservation, and biogeography. Here, we describe the structure and extension of the HERBase database, compiled from projects and research teams that have been dedicated to study herb ecology in Amazonia.

MATERIAL AND METHODS

Definitions

Defining an herb is not straightforward, although dictionaries define herbs as plants without a woody stem. For example, the Merriam-Webster dictionary defines herbs as seed-producing annual, biennial, or perennial plants that do not develop persistent woody tissue but die down at the end of a growing season. The term "seed-producing" excludes ferns and lycophytes. And, in practice, defining woodiness may be problematic. For example, some plants are classified as subshrubs by some researchers, but as herbs by others, and some monocotyledons (such as palms, and several Zingiberales) and ferns have fibrous layers around pseudostems or petioles that appear woody, although technically these groups do not produce true wood.

In practice, researchers have tended to consider all ferns, lycophytes, and monocotyledons as part of the tropical forest

herb community, but not to include non-monocotyledonous angiosperms plants that might fall in the ambiguous herb or shrub or subshrub categories (Poulsen and Balslev 1991). This issue complicates analyses based on data compiled from several sources since most often the definition of what are herbs is not explicit in each particular study. There is an obvious need to use clear definitions and standardization in future studies (see the section on recommended best practices in the Supplementary Material, Appendix S1).

Another methodological issue that may create differences among datasets is whether all possible substrates have been included in the sampling. Some herb species are obligate terrestrials and others are obligate epiphytes, but there are also several intermediate habits. For example, many species can grow both terrestrially and as epiphytes, and for species with a climbing habit, it may be difficult to determine whether they have a ground connection or not. Moreover, the substrate may change during the lifetime of an individual plant: an originally terrestrial individual may climb up a tree trunk and later lose its ground connection and become an epiphyte, or an originally epiphytic individual may create a ground connection as it grows larger. Many studies on ferns and lycophytes have taken into account all individuals, including epiphytes and climbers on the lower parts of tree trunks (Tuomisto *et al.* 2003a, 2003b, 2003c, 2016; Higgins *et al.* 2011). However, other studies have only included terrestrial individuals (Poulsen and Balslev 1991) or species that are known mostly to be terrestrial (Tuomisto and Poulsen 1996, 2000; Zuquim *et al.* 2012, 2014; Moulatlet *et al.* 2014; Tuomisto *et al.* 2019). Studies are not always clear on the definition of the life forms or habits and substrates included, and the simple assignment of a species to a substrate may be inaccurate. Future studies should pay special attention to clear definitions and documentation of substrates to enhance the usefulness of the data.

Data compilation

In this first compilation, we only considered herb plots located within the limits of the Amazon basin, according to the concept of Amazonia *sensu latissimo* proposed by Eva and Huber (2005) (Figure 1), which includes areas of the areas of savanna in the Cerrado Biome as well as montane areas that drain into the Amazon River. This delimitation of Amazonia encompasses an area of 7,595,000 km² and includes areas from Brazil, Bolivia, Peru, Ecuador, Colombia, Venezuela, Guyana, Suriname, and French Guiana. The studies from which data initially have been compiled into HERBase have had various objectives: complete floristic inventory (Duivenvoorden 1995), assessment of the floristic composition in specific forest types (van Andel 2003; Linares-Palomino *et al.* 2013), determinants of species richness and/or composition at a local scale (Tuomisto and Ruokolainen 1994; Costa *et al.* 2005; Drucker *et al.* 2008; Magalhães and Lopes 2015; da Silva *et al.*

2021; Rodrigues *et al.* 2021) or at a regional scale (Tuomisto and Poulsen, 1996; Tuomisto *et al.* 2003a, 2014, 2016; Zuquim *et al.* 2012, 2014; Figueiredo *et al.* 2014; Moulatlet *et al.* 2014; Riaño and Moulatlet 2022), effects of anthropogenic disturbance on herb assemblages (Costa and Magnusson 2002; de Polari Alverga *et al.* 2021) and the effects of past human forest modifications on herb composition (Quintero-Vallejo *et al.* 2015). However, future contributions to HERBase need not be restricted to these types of studies and the database is intended as a data repository for voluntary deployment of data by accessing <https://www.gov.br/inpa/pt-br/projetos/herbase>.

Data curation

Taxonomic standardization is fundamental when compiling vegetation databases to address nomenclature redundancy caused by the multitude of synonyms characterizing botanical literature (Kalwij 2012), and to perform comparative analyses across data sets. In HERBase, all data compiled or received is standardized to a uniform taxonomy based on the Flora and Fungi of Brazil (<http://floradobrasil.jbrj.gov.br>; BFG 2021, 2018), first, with the use of the 'flora' package (<https://github.com/gustavobio/flora>) in the R environment (R Core Team 2022) and then by consulting specialized taxonomic works and/or specialist taxonomists. Also, members from HERBase dedicated to the taxonomy of specific plant groups (e.g., ferns

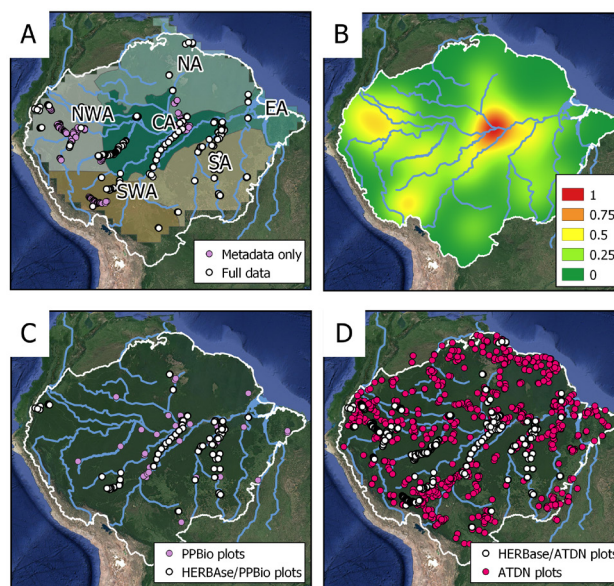


Figure 1. Location of HERBase sampling plots included by June 2022: A – within geographic regions, as defined by Feldpausch *et al.* (2012). White dots represent plots with data included in the database; violet dots represent plots with only metadata available in the database; B – Sampling density, according to the Kernel estimation, varies from 0 to 1, as color-coded from dark green (0) to red (1); C – in relation to sampling plots of the PPBio program (<https://ppbio.inpa.gov.br/>). Violet dots represent plots with complete data sets included in HERBase; white dots represent plots with only metadata available in HERBase; D – in relation to ATDN tree-plot network (<https://www.atdn.myspecies.info>). White dots indicate plots with complete data included in HERBase; pink dots represent the distribution and coverage of tree inventories. This figure is in color in the electronic version.

and lycophytes) are responsible for curating and updating data information before it enters the database.

Sampling coverage

Based on the coordinates of the plots compiled in the metadata, we calculated the spatial density of plots using the Kernel density estimation implemented in QGIS. We also addressed the geographical plot coverage using a classification of the Amazon basin as defined by Feldpausch *et al.* (2012) [northwestern Amazonia (NWA); southwestern Amazonia (SWA); southern Amazonia (SA); central Amazonia (CA); Guiana Shield (GS); and eastern Amazonia (EA)].

RESULTS

Database characteristics

As of June 2022, HERBase had 1381 plots with inventory data included, and 342 plots with only metadata included and data available upon request to the principal investigator. The inventory of herbs can be based on different metrics to provide an estimate of abundance (e.g., direct counting of individuals, estimates of cover, frequency of occupation). In HERBase, there is a predominance of density data based on the counting of individuals (89% of the plots). Cover data is available for 23% of the plots, frequently in combination with counting data (12.3% of total data).

The most common sampling unit in the herb inventories was a fixed-area plot. Sampling designs without a defined area, such as distance-sampling (Buckland *et al.* 2005) are rarely used for herbs and are not present in HERBase up to now. The size of the included plots varies widely (Figure 2). The most common plot areas range between 500 m² and 1000 m² (48%), followed by larger plots of ≥ 1000 m² (29%), and plots smaller than 500 m² (23%). Plot shape varies from square (8.3%) to rectangular transects (40.6%), to transects that adjust to the terrain altitudinal contour and plot width to the organism sampled (51.1%). The largest fraction of plots included in HERBase contains inventory data for all terrestrial herbs (35%), followed by inventories of only ferns and lycophytes (30%). The remaining data are quite variable, including inventories of ferns and lycophytes + Zingiberales (12%), all terrestrial herbs + epiphytes (10%), only Zingiberales (5%),

ferns and lycophytes + monocotyledons (3%) and ferns and lycophytes + Araceae + Marantaceae (2%) (Figure 2).

The distribution of plots in relation to local landscape heterogeneity varied according to the aims of the original study. The two most common sampling-unit types in HERBase are: 1) 5-m wide transects (mostly ≥ 500 m long) that run across the local topographical variation (25% of included plots); and 2) 2-m wide and 250-m long plots that maintain a fixed position along elevational contour lines, belonging to PPBio infrastructure (51%). The first design aims to produce data that are representative of the landscape as a whole by maximising the hydrological variation within the transect, thus increasing the diversity of species within the transect and facilitating regional comparisons (Tuomisto *et al.* 2003c). The second design aims to produce data that are representative of local hydrological or soil conditions by minimizing the topographical variation within each plot, and the landscape-scale variation is captured by establishing many plots per site (Magnusson *et al.* 2005).

Sampling coverage

The plots are unevenly distributed across geographical regions, with 37 % located in central Amazonia, 24.7% in northwestern Amazonia, 15.1% in southwestern Amazonia, 14% in southern Amazonia, 6% in northern Amazonia and 2% in eastern Amazonia (Figure 1a). The highest density of sampling plots (Figure 1b) is located in central Brazilian Amazonia (in the surroundings of Manaus), with other sampling clusters in northwestern Amazonia (around the Peruvian city of Iquitos) and in southwestern Amazonia, close to the border between Brazil and Bolivia and along the Madre de Dios River (Peru). In general, there is a higher density of plots along the main rivers compared to inland areas.

When compared with other plot networks in Amazonia, particularly PPBio and ATDN, HERBase has a much more restricted distribution. Many of the HERBase plots belong to the PPBio plot network (white dots in Figure 1c), while PPBio plot in which herbs were not as yet sampled are represented by violet dots (Figure 1c). The ATDN network has a broad and dense coverage of tree sampling plots over the whole Amazon region, and basically all the PPBio plots included in the ATDN have also been sampled for herbs (Figure 1d). Some

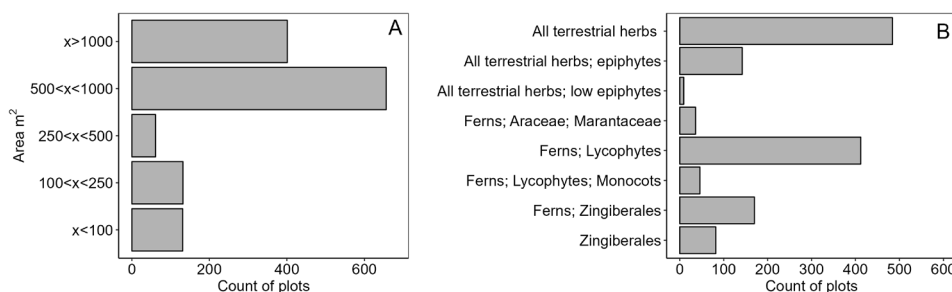


Figure 2. Frequency of herb sampling plots included in HERBase until June 2022 according to area (m²) (A) and group(s) of herbs sampled (B).

of these plots have herb data that have not yet been included in HERBase. Most of the ATDN plots with HERBase data in Brazil coincide with PPBio plots and this spatial integration of sampling plots allows the integration of scientific data.

DISCUSSION

The HERBase initiative has managed to compile extensive plot-based information on ground herbs sampled in all Amazonian geographic regions. To the best of our knowledge, the >1,700 inventory plots included in HERBase represent a significant part of understory herb community inventories implemented in Amazonia.

Sampling coverage

Our data compilation showed that the surroundings of the large urban centres in Amazonia have the highest plot density. Although many efforts to sample remote areas have been undertaken in the past decades, sampling is still constrained by complex logistics and high costs, thus being biased towards the areas that are most accessible from urban centers by road or river. Similar access constraints to sampling areas have affected inventories of Amazonian trees (Nelson *et al.* 1990; Hopkins 2007) and animals (Oliveira *et al.* 2016b).

In the HERBase data, the surroundings of Manaus (Brazil) stand out as especially intensively collected, followed by the surroundings of Iquitos (Peru). Both of these areas are of interest to researchers also due to their high geodiversity (Higgins *et al.* 2011; Figueiredo *et al.* 2014). Smaller concentrations of sampling are found along the river Madre de Dios in Peru, and the rivers Juruá and Tapajós in Brazil. The fact that most of our data come from central and northwestern Amazonia reflects the long-term efforts of a few research groups in local research institutions. Large sampling gaps remain in other Amazonian regions, including, for example, the region of the large urban area of Belém (Brazil), which so far has not attracted researchers to do herb inventories. The comprehensive sampling of such regions would not only improve our understanding of Amazonian herbs, but also provide environmental description of these regions, as basic environmental data are usually collected together with floristic inventories. In addition, herb inventory data can be used to infer and map environmental variables across Amazonia even when direct environmental measurements are not available (Zuquim *et al.* 2019; Tuomisto *et al.* 2019). There is great potential in the use of standardized sampling plots such as those used by the PPBio and RAINFOR projects over broad spatial scales, as it decreases the costs of infrastructure implementation and allows direct comparison of biodiversity data among sampling sites.

HERBase functioning

HERBase emerged from personal contacts among researchers interested in herbs, who agreed to share data and

metadata from their plots. Taxonomic data are curated with the most updated sources and a committee of taxonomists decides on ambiguous cases. Metadata on plot location, size, shape, forest type, and minimum plant sampling size and habit is included for all datasets. HERBase is based on the principles of equality among partners, where all contributors have the same rights to propose uses for the full dataset or parts of it. A five-member HERBase Management Committee is elected among participating researchers of the network, two of which are in a coordinating role, in addition to two substitutes. Every two years, at least one new member becomes part of the committee, replacing the participating researcher with the longest time on the committee or the one(s) who wishes to resign. The Management Committee meets at the request of any of the participating members and/or by invitation of the coordinators, to discuss data use requests and to plan events, scientific dissemination, projects, and publications, among others.

The participants have priority in data use, and any of them can request data for specific uses. Most (but not all) of the data now compiled in HERBase have been made available in connection with already published articles. The advantages of participating in HERBase and requesting data internally are that a) these data have already passed through taxonomic standardization; b) the management of the database aims to minimize overlap among project proposals that would address similar research questions; and c) HERBase makes sure that the data owners are properly consulted in advance and allows them to decide whether the data will be used for the planned purpose or not, giving proper opportunities of authorship to all participants.

On the website dedicated to HERBase (www.gov.br/inpa/pt-br/projetos/herbase), the associated metadata are made openly available for anyone to explore. If interest in using the data emerges from this, requests for data use will be evaluated by the management committee and the data owners will be consulted. Detailed information on how to contribute to HERBase, as well as specific data requirements, can be found on the dedicated website. HERBase welcomes all datasets dedicated to the study of herbs from vegetation plots located within the limits of the Amazon biome as defined here. HERBase aims to contribute to the understanding of herb diversity in general with a broad biogeographic focus, so all types of data are welcome.

CONCLUSIONS

Although HERBase represents an important first step to organizing Amazonian herb inventories, we recommend that future efforts in sampling herbs consider the current biases in plot location in Amazonia. The lack of data from areas in the west and east of the region, where other plant groups have been much more intensively sampled precludes

a general understanding of herb species diversity and its relationship with other components of biodiversity. We also strongly recommend that new sampling efforts aim at using standardized sampling methods and following the best practices outlined in Supplementary Material, Appendix S1. HERBase is an effort to integrate and fill gaps in the knowledge about the distribution of herb species in Amazonia, and we hope it will encourage more studies of understory herbs. We encourage colleagues in possession of herb data to join the HERBase initiative.

ACKNOWLEDGMENTS

Funding from Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (Brazil) (proc. # 423574/2018-3 to TA, 317091/2021-2 and 441443/2016-8 to TEA, 472799/03-7 to DPD and 47890820/2-1 to FRCC) is acknowledged. GMM acknowledges the postdoctoral grant from Consejo Nacional de Ciencia y Tecnología (Mexico) project SEP-CONACYT CB-2017-2018 (A1-S-34563). RLP acknowledges funds from various projects with GeoPark Perú, Transportadora de Gas del Perú, Hunt Oil Company Peru and EcoPetrol Perú (Peru). AGM acknowledges Fundação de Amparo à Pesquisa de Rondônia (FAPERO) (Brazil) in the calls FAPERO/Universal 2018 and FAPERO/PRO-Rondônia. ECP acknowledges the Centro de Pesquisa do Pantanal (Brazil). EQV acknowledges Instituto Boliviano de Investigación Forestal, IBIF (Bolivia) and INREF (Holanda). DST acknowledges Reserva Biológica do Uatumã (Brazil). HT acknowledges many colleagues for collaboration in the field, especially Kalle Ruokolainen, Glenda Cárdenas, Adriana Zegarra and Mark Higgins, and several funding agencies, especially the Academy of Finland (most recently grants # 273737 and 344733). FRCC acknowledges the Instituto Nacional de Pesquisas da Amazônia (INPA), Programa de Graduação em Ecologia do INPA, Programa de Pesquisas em Biodiversidade (PPBio), Programa de Pesquisas Ecológicas de Longa Duração (PELD), Projeto Dinâmica Biológica de Fragmentos Florestais (PDBFF), Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio), and Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis (IBAMA) (Brazil) for logistical support, and PPBio for funding. Fernando Figueiredo is greatly acknowledged for sharing and curating data. We are all thankful to numerous field assistants for contributing to data collection over many years, especially João Batista, José da S. Lopes, José T. do Nascimento, José da Silva and Wellington da Silva.

REFERENCES

- van Andel, T.R. 2003. Floristic composition and diversity of three swamp forests in northwest Guyana. *Plant Ecology*, 167: 293–317.
- Beck, J.J. 2021. Variation in plant–soil interactions among temperate forest herbs. *Plant Ecology*, 222: 1225–1238.
- Buckland, S.T.; Anderson, D.R.; Burnham, K.P.; Laake, J.L. 2005. Distance Sampling. In: Armitage, P; Colton, T. (Ed.). *Encyclopedia of Biostatistics*, John Wiley & Sons, Ltd.
- Cicuzza, D.; Krömer, T.; Poulsen, A.D.; Abrahamczyk, S.; Delhotal, T.; Piedra, H.M.; *et al.* 2013. A transcontinental comparison of the diversity and composition of tropical forest understory herb assemblages. *Biodiversity and Conservation*, 22: 755–772.
- Connell, J.H. 1971. On the role of the natural enemies in preventing competitive exclusion in some marine animals and in rain forest trees. In: Den Boer, P.J.; Gradwell, G.R. (Ed.). *Dynamics of Populations*, Center for Agricultural Publishing and Documentation, Wageningen, p.298–312.
- Cornwell, W.K.; Pearse, W.D.; Dalrymple, R.L.; Zanne, A.E. 2019. What we (don't) know about global plant diversity. *Ecography*, 42: 1819–1831.
- Costa, F.R.C.; Magnusson, W.E. 2002. Selective logging effects on abundance, diversity, and composition of tropical understory herbs. *Ecological Applications*, 12: 807–819.
- Costa, F.R.C.; Magnusson, W.E.; Luizao, R.C. 2005. Mesoscale distribution patterns of Amazonian understory herbs in relation to topography, soil and watersheds. *Journal of Ecology*, 93: 863–878.
- Diniz-Filho, J.A.F.; Terribile, L.C.; Da Cruz, M.J.R.; Vieira, L.C.G. 2010. Hidden patterns of phylogenetic non-stationarity overwhelm comparative analyses of niche conservatism and divergence: Non-stationarity in niche evolution. *Global Ecology and Biogeography*, 19: 916–926.
- Dodson, C.H.; Gentry, A.H. 1978. Flora of the Rio Palenque Science Center: Los Rios Province, Ecuador. *Selbyana*, 4: 1–628.
- Draper, F.C.; Costa, F.R.C.; Arellano, G.; Phillips, O.L.; Duque, A.; Macía, M.J.; *et al.* 2021. Amazon tree dominance across forest strata. *Nature Ecology & Evolution*, 5: 757–767.
- Drucker, D.P.; Costa, F.R.C.; Magnusson, W.E. 2008. How wide is the riparian zone of small streams in tropical forests? A test with terrestrial herbs. *Journal of Tropical Ecology*, 24: 65–74.
- Duivenvoorden, J.F. 1995. Tree species composition and rain forest–environment relationships in the middle Caquetá area, Colombia, NW Amazonia. *Vegetatio*, 120: 91–113.
- Eva, H.D.; Huber, O. 2005. A proposal for defining the geographical boundaries of Amazonia. (<http://library.wur.nl/WebQuery/wurpubs/fulltext/26674>). Accessed on 08 Aug 2017.
- Feldpausch, T.R.; Lloyd, J.; Lewis, S.L.; Brien, R.J.W.; Gloor, M.; Monteagudo Mendoza, A.; *et al.* 2012. Tree height integrated into pantropical forest biomass estimates. *Biogeosciences*, 9: 3381–3403.
- Figueiredo, F.O.G.; Costa, F.R.C.; Nelson, B.W.; Pimentel, T.P. 2014. Validating forest types based on geological and land-form features in central Amazonia. *Journal of Vegetation Science*, 25: 198–212.
- Figueiredo, F.O.G.; André, T.; Moullet, G.M.; Saka, M.N.; Araujo, M.H.T.; Tuomisto, H.; *et al.* 2022. Linking high diversification rates of rapidly growing Amazonian plants to geophysical landscape transformations promoted by Andean uplift. *Botanical Journal of the Linnean Society*, 199: 36–52.

- Gentry, A.H. 1988. Changes in plant community diversity and floristic composition on environmental and geographical gradients. *Annals of the Missouri Botanical Garden*, 75: 1–34.
- George, L. O.; Bazzaz, F. A. 1999. The fern understory as an ecological filter: emergence and establishment of canopy-tree seedlings. *Ecology*, 80: 833–845.
- Grand, J.; Cummings, M.P.; Rebelo, T.G.; Ricketts, T.H.; Neel, M.C. 2007. Biased data reduce efficiency and effectiveness of conservation reserve networks. *Ecology Letters*, 10: 364–374.
- Higgins, M.A.; Ruokolainen, K.; Tuomisto, H.; Llerena, N.; Cardenas, G.; Phillips, O.L.; et al. 2011. Geological control of floristic composition in Amazonian forests. *Journal of Biogeography*, 38: 2136–2149.
- Hopkins, M.J.G. 2007. Modelling the known and unknown plant biodiversity of the Amazon Basin. *Journal of Biogeography*, 34: 1400–1411.
- Hortal, J.; Bello, F. de; Diniz-Filho, J.A.F.; Lewinsohn, T.M.; Lobo, J.M.; Ladle, R.J. 2015. Seven shortfalls that beset large-scale knowledge of biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, 46: 523–549.
- Hubbell, S. 2001. *The Unified Neutral Theory of Biodiversity and Biogeography*, 1st ed. Princeton University Press, New Jersey, 392p.
- Janzen, D.H. 1970. Herbivores and the number of tree species in tropical forests. *The American Naturalist*, 104: 501–528.
- Jones, M.M.; Tuomisto, H.; Olivares, P.C. 2008. Differences in the degree of environmental control on large and small tropical plants: just a sampling effect? *Journal of Ecology*, 96: 367–377.
- Jones, M.M.; Ferrier, S.; Condit, R.; Manion, G.; Aguilar, S.; Pérez, R. 2013. Strong congruence in tree and fern community turnover in response to soils and climate in central Panama. *Journal of Ecology*, 101: 506–516.
- Kalwij, J.M. 2012. Review of ‘The Plant List, a working list of all plant species.’ *Journal of Vegetation Science*, 23: 998–1002.
- Linares-Palomino, R.; Chávez, G.; Pérez, E.; Goshima, F.; Zamora, H.; Deichmann, J.; et al. 2013. Patrones de diversidad y composición en comunidades de pteridophyta, aves, anfibios, reptiles y murciélagos en la cuenca del río Tapiche, Loreto. In: Linares-Palomino, R.; Deichmann, J.; Alonso, A. (Ed.). *Biodiversidad y Uso de Recursos Naturales en la Cuenca Baja del Río Tapiche, Loreto, Perú*. Instituto de Investigación de la Amazonía Peruana, Peru, p.14–55.
- Magalhães, J.L.L.; Lopes, M.A. 2015. Species richness and abundance of low-trunk herb epiphytes in relation to host tree size and bark type, eastern Amazonia. *Revista Árvore*, 39: 457–466.
- Magnusson, W.E. 2013. *Biodiversity and Integrated Environmental Monitoring: the RAPELD system in the Amazon*. 1st ed. Attema Editorial, Santo André, 351p.
- Magnusson, W.E.; Lima, A.P.; Luizão, R.; Luizão, F.; Costa, F.R.C.; Castilho, C.V. de; et al. 2005. RAPELD: a modification of the Gentry method for biodiversity surveys in long-term ecological research sites. *Biota Neotropica*, 5: 19–24.
- Magnusson, W.E. 2019. Biodiversity: the chasm between what we know and we need to know. *Anais da Academia Brasileira de Ciências*, 91:1–3.
- Moerman, D.E.; Estabrook, G.F. 2006. The botanist effect: counties with maximal species richness tend to be home to universities and botanists. *Journal of Biogeography*, 33: 1969–1974.
- Moulatlet, G.M.; Costa, F.R.C.; Rennó, C.D.; Emilio, T.; Schiatti, J. 2014. Local hydrological conditions explain floristic composition in lowland Amazonian forests. *Biotropica*, 46: 395–403.
- Moulatlet, G.M.; Zuquim, G.; Figueiredo, F.O.G.; Lehtonen, S.; Emilio, T.; Ruokolainen, K.; et al. 2017. Using digital soil maps to infer edaphic affinities of plant species in Amazonia: Problems and prospects. *Ecology and Evolution*, 7: 8463–8477.
- Nelson, B.W.; Ferreira, C.A.C.; da Silva, M.F.; Kawasaki, M.L. 1990. Endemism centres, refugia and botanical collection density in Brazilian Amazonia. *Nature*, 345: 714–716.
- Oliveira, B.F.; Machac, A.; Costa, G.C.; Brooks, T.M.; Davidson, A.D.; Rondinini, C.; et al. 2016a. Species and functional diversity accumulate differently in mammals. *Global Ecology and Biogeography*, 25: 1119–1130.
- Oliveira, U.; Paglia, A.P.; Brescovit, A.D.; de Carvalho, C.J.B.; Silva, D.P.; Rezende, D.T.; et al. 2016b. The strong influence of collection bias on biodiversity knowledge shortfalls of Brazilian terrestrial biodiversity. *Diversity and Distributions*, 22: 1232–1244.
- Paiva, D.N.A.; Perdiz, R. de O.; Almeida, T.E. 2021. Using near-infrared spectroscopy to discriminate closely related species: a case study of neotropical ferns. *Journal of Plant Research*, 134: 509–520.
- Perea, R.; Schroeder, J.W.; Dirzo, R. 2022. The herbaceous understory plant community in the context of the overstory: An overlooked component of tropical diversity. *Diversity*, 14: 800. doi.org/10.3390/d14100800
- de Polari Alverga, P.P.; Miranda, P.N.; da Silva Oliveira, R.; Morato, E.F. 2021. Effects of forest succession on the richness and composition of Zingiberales in a forest fragment in the southwestern Amazon. *Brazilian Journal of Botany*, 44: 491–502.
- Poulsen, A.D.; Balslev, H. 1991. Abundance and cover of ground herbs in an Amazonian rain forest. *Journal of Vegetation Science*, 2: 315–322.
- Quintero-Vallejo, E.; Klomberg, Y.; Bongers, F.; Poorter, L.; Toledo, M.; Peña-Claros, M. 2015. Amazonian dark earth shapes the understory plant community in a Bolivian forest. *Biotropica*, 47: 152–161.
- R Core Team. 2022. *R: A language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria. (<https://www.r-project.org/>). Accessed on 28 Feb 2019.
- Riaño, K.; Moulatlet, G.M. 2022. Floristic and functional diversity of ferns and lycophytes at three elevational zones in the eastern slopes of the northern Andes, Ecuador. *Acta Amazonica*, 52: 149–157.
- Ribas, C.C.; Gaban-Lima, R.; Miyaki, C.Y.; Cracraft, J. 2005. Historical biogeography and diversification within the Neotropical parrot genus *Pionopsitta* (Aves: Psittacidae). *Journal of Biogeography*, 32: 1409–1427.
- Ribeiro, J.E.L.S.; Hopkins, M.J.G.; Vicentini, A.; Sothers, C.A.; Costa, M.A.S.; Brito, J.M.; et al. 1999. *Flora da Reserva Ducke*:

- Guia de Identificação de Plantas Vasculares de uma Floresta de Terra-Firme na Amazônia Central*. Editora INPA, Manaus, 800p.
- Rodrigues, D.B.; Oliveira, M.H.V. de; Silva, A. da C.; Almeida, T.E.; André, T.; Mortati, A.F. 2021. Ground-herb communities of *terra firme* riparian forests of the lower Tapajós River in the Brazilian Amazon. *Rodriguésia*, 72: e00052020.
- Royo, A.A.; Carson, W.P. 2006. On the formation of dense understory layers in forests worldwide: consequences and implications for forest dynamics, biodiversity, and succession. *Canadian Journal of Forest Research*, 36: 1345–1362.
- Ruokolainen, K.; Linna, A.; Tuomisto, H. 1997. Use of Melastomataceae and pteridophytes for revealing phytogeographical patterns in Amazonian rain forests. *Journal of Tropical Ecology*, 13: 243–256.
- Ruokolainen, K.; Tuomisto, H.; Macía, M.J.; Higgins, M.A.; Yli-Halla, M. 2007. Are floristic and edaphic patterns in Amazonian rain forests congruent for trees, pteridophytes and Melastomataceae? *Journal of Tropical Ecology*, 23: 13–25.
- da Silva, J.G.; Vieira, T.B.; Mews, H.A. 2021. Fine-scale effect of environmental variation and distance from watercourses on pteridophyte assemblage structure in the western Amazon. *Folia Geobotanica*, 56: 69–80.
- ter Steege, H.; Pitman, N.; Sabatier, D.; Castellanos, H.; van der Hout, P.; Daly, D.C.; *et al.* 2003. A spatial model of tree α -diversity and tree density for the Amazon. *Biodiversity & Conservation*, 12: 2255–2277.
- ter Steege, H.; Pitman, N.C.A.; Phillips, O.L.; Chave, J.; Sabatier, D.; Duque, A.; *et al.* 2006. Continental-scale patterns of canopy tree composition and function across Amazonia. *Nature*, 443: 444–447.
- ter Steege, H.; Pitman, N.C.A.; Sabatier, D.; Baraloto, C.; Salomão, R.P.; Guevara, J.E.; *et al.* 2013. Hyperdominance in the Amazonian tree flora. *Science*, 342: 1243092.
- Tuomisto, H.; Poulsen, A.D. 1996. Influence of edaphic specialization on pteridophyte distribution in neotropical rain forests. *Journal of Biogeography*, 23: 283–293.
- Tuomisto, H.; Poulsen, A.D. 2000. Pteridophyte diversity and species composition in four Amazonian rain forests. *Journal of Vegetation Science*, 11: 383–396.
- Tuomisto, H.; Ruokolainen, K. 1994. Distribution of Pteridophyta and Melastomataceae along an edaphic gradient in an Amazonian rain forest. *Journal of Vegetation Science*, 5: 25–34.
- Tuomisto, H.; Ruokolainen, K.; Yli-Halla, M. 2003a. Dispersal, environment, and floristic variation of western Amazonian forests. *Science*, 299: 241–244.
- Tuomisto, H.; Zuquim, G.; Cárdenas, G. 2014. Species richness and diversity along edaphic and climatic gradients in Amazonia. *Ecography*, 37: 1034–1046.
- Tuomisto, H.; Ruokolainen, K.; Aguilar, M.; Sarmiento, A. 2003b. Floristic patterns along a 43-km long transect in an Amazonian rain forest. *Journal of Ecology*, 91: 743–756.
- Tuomisto, H.; Poulsen, A.D.; Ruokolainen, K.; Moran, R.C.; Quintana, C.; Celi, J.; *et al.* 2003c. Linking floristic patterns with soil heterogeneity and satellite imagery in Ecuadorian Amazonia. *Ecological Applications*, 13: 352–371.
- Tuomisto, H.; Moulatlet, G.M.; Balslev, H.; Emilio, T.; Figueiredo, F.O.G.; Pedersen, D.; *et al.* 2016. A compositional turnover zone of biogeographical magnitude within lowland Amazonia. *Journal of Biogeography*, 43: 2400–2411.
- Tuomisto, H.; Van Doninck, J.; Ruokolainen, K.; Moulatlet, G.M.; Figueiredo, F.O.G.; Sirén, A.; *et al.* 2019. Discovering floristic and geoecological gradients across Amazonia. *Journal of Biogeography*, 46: 1734–1748.
- Vormisto, J.; Phillips, O.L.; Ruokolainen, K.; Tuomisto, H.; Vásquez, R. 2000. A comparison of fine-scale distribution patterns of four plant groups in an Amazonian rainforest. *Ecography*, 23: 349–359.
- Whittaker, R.J.; Araújo, M.B.; Jepson, P.; Ladle, R.J.; Watson, J.E.M.; Willis, K.J. 2005. Conservation Biogeography: assessment and prospect. *Diversity and Distributions*, 11: 3–23.
- Yang, W.; Ma, K.; Kreft, H. 2013. Geographical sampling bias in a large distributional database and its effects on species richness-environment models. *Journal of Biogeography*, 40: 1415–1426.
- Zuquim, G.; Tuomisto, H.; Costa, F.R.C.; Prado, J.; Magnusson, W.E.; Pimentel, T.; *et al.* 2012. Broad scale distribution of ferns and lycophytes along environmental gradients in Central and Northern Amazonia, Brazil. *Biotropica*, 44: 752–762.
- Zuquim, G.; Tuomisto, H.; Jones, M.M.; Prado, J.; Figueiredo, F.O.G.; Moulatlet, G.M.; *et al.* 2014. Predicting environmental gradients with fern species composition in Brazilian Amazonia. *Journal of Vegetation Science*, 25: 1195–1207.
- Zuquim, G.; Stropp, J.; Moulatlet, G.M.; Van doninck, J.; Quesada, C.A.; Figueiredo, F.O.G.; *et al.* 2019. Making the most of scarce data: Mapping soil gradients in data-poor areas using species occurrence records. *Methods in Ecology and Evolution*, 10: 788–801.

RECEIVED: 14/11/2022

ACCEPTED: 31/01/2023

ASSOCIATE EDITOR: Natalia Ivanauskas



This is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

SUPPLEMENTARY MATERIAL (only available in the electronic version)André *et al.* HERBase: A collection of understory herb vegetation plots from Amazonia**Appendix S1. Protocol for best practice in the sampling of understory herbs****1) To ensure adequate ecological analysis based on plot data.**

The more accurate the information about the plots (i.e., the metadata), the better the representation of environmental conditions, both *in situ* and extracted from remote sensing. We recommend to:

- (a) record geographic coordinates as accurately as possible at different points of the sampling plot;
- (b) record time and date, participants (researchers, students and field assistants), compass direction (if your transect is linear rather than square);
- (c) whenever possible, take pictures of the plot and its environmental features;
- (d) make a description of the environmental characteristics of each plot, as far as possible in relation to all factors that may affect the abundance and composition of the herbs. As much as possible, obtain quantitative measurements of the elements that affect plants (water, light and nutrients). The following are suggested: altitude; topographic position of the plot; distance from water sources such as rivers, streams, lakes and puddles; vegetation structure; presence/intensity of anthropic disturbances; soil texture and concentrations of elements that are known to be plant nutrients; light availability.
- (e) preferably, make soil collections at different points of each plot. If soil analysis cannot be done immediately, store the soil in a dry place away from direct light exposure after drying.

2) To ensure standardized abundance comparisons.

Integrative analyses demand standardized data, and the quality of the analyses is directly dependent on this standardization. We recommend to:

- (a) clearly define the minimum size limit of the sampled plants;
- (b) clearly define what the individual is (in the case of counting), and in the case of clonal species, whether to count the clones (genets) or ramets as individuals. If you are counting clones as individuals, define the extent of the patch or the maximum distance between ramets that corresponds to one individual;
- (c) clearly define which plant life forms or habits are included in the studied community (e.g., obligate terrestrial, facultative terrestrial, hemiepiphyte, epiphyte);
- (d) take note in the field of the habits of individuals, especially in the case of species that can assume different habits;

(e) in the case of epiphytes or hemi-epiphytes, clearly define the minimum and maximum sampling heights on the phorophytes.

3) To ensure standardized comparison of species composition and richness. We recommend to:

- (a) collect samples of species and morphotypes in each plot. If plots are used for permanent monitoring and individuals cannot be collected within the plot, individuals of the same morphotype should be searched in the immediate vicinity outside the plot, and photos of individuals within the plot should be used for documentation;
- (b) for each plot, make vouchers of at least one adult mature individual of each sampled morphotype or species recorded;
- (c) for small individuals of ferns and lycophytes, collect the complete plant, including the rhizome. To avoid killing large individuals, sampling can be restricted to leaves only, but then the rhizome type should be documented by photographing or describing in the specimen metadata;
- (d) document the variability of vegetative morphological characteristics of each species within each plot, at least with photos. Many important groups of Amazonian terrestrial herbs can be identified by experts from good photographs of the specimens. Take at least one photo of the complete individual, a branch with at least one full leaf, and any fertile parts that may be present (flowers, fruit, sori). We recommend depositing these georeferenced images in taxonomic databases;
- (e) record, as much as possible, the size (e.g., height) of individuals;
- (f) for flowering plants, when the plant is fertile, preferably collect flowers and/or inflorescences in a wet way (preferably 70 parts of ethanol, 27 parts of water, and 3 parts of glycerin) and deposit them under the same voucher number of the dried pressed voucher in the herbarium. This wet collection preserves the three-dimensional structure of the reproductive parts, which is important for the determination of most taxonomic groups;
- (g) clearly record the bibliographic sources, names of specialist botanists, reference collections, and classification system used for species identification;
- (h) sample the entire herb community, to the greatest extent possible, rather than only taxonomic subgroups. When subgroups are sampled, clearly document what they are;
- (i) preserve samples in silica for genetic analysis

Further information on materials to take to the field and how to collect samples on environmental variables can be found in website of the Research Program on Biodiversity

(*Programa de Pesquisa em Biodiversidade – PPBio*) (<https://ppbio.inpa.gov.br/en/methods>).

4) Sampling protocol suggestion.

Many types of sampling designs and ways of measuring the occurrence or abundance of herbs exist, and different methods have been used by different researchers to answer the same or different questions. However, this diversity can become a problem when trying to combine datasets obtained with different methods, especially in large-scale integrative analyses. Thus, as a step towards better future data integration, we suggest a sampling protocol and discuss the reasons for adopting it, as well as possible adjustments so that its application is viable in different environments.

At least one representative voucher specimen of each species or morphospecies rooted in the plot should be collected, to assure the existence of testimony material for identification confirmation and other kinds of studies. Many contemporary techniques can be applied for taxonomic identification of sterile material, such as FT-NIR spectroscopy, which works well for both angiosperm (e.g. Paiva *et al.* 2021) and ferns (G. Moulatlet pers. info). We obtained 16 spectral readings per individual from the adaxial and abaxial surfaces of 100 specimens belonging to 13 species. The analyses included all 1557 spectral variables. We tested different datasets (adaxial + abaxial, adaxial, and abaxial. Although herbaria generally prefer to receive fertile material, sterile specimens are usually accepted if they were collected in permanent plots and come with good enough metadata. Whenever possible, the entire plant should be photographed in the field, to record potentially important details for identification, such as habit, fertile organs, and any coloured parts. It is important to write down the camera file name in the field and relate it to the specimen registration number. The collected material

must be labelled while still in the field and the registration number included in the field sheet. Collections should be kept in closed plastic bags to prevent plants from wilting before being pressed. Whenever possible, it is recommended to complement the collection made inside the plot with the collection of a whole plant of the same species outside the plot. However, in case of uncertainty in comparing morphotypes, make sure that complementary collections have a different number, to avoid mixed collections.

A complete count of terrestrial herb specimens ≥ 5 cm tall should ideally be carried out in each segment of the plot. The size criterion is a recommendation, as the inventory data compiled in HERBase include different height criteria. For ferns, the UTU protocol uses a minimum leaf length of 10 cm, which excludes fern gametophytes and small juveniles of the sporophytes, as these can be both numerous and difficult to identify. It is important to make sure to only include plants rooted within the established plot width. If in doubt whether a particular specimen is an obligatory terrestrial herb or is only temporarily in the herbaceous stratum, collect it and record the status of the individual. Later, with a reliable determination of your samples, a decision to keep or exclude the individual plant from the sample can be made.

Regarding the type of plot, we recommend that users choose one of the most common types already presented in HERBase. For complete herb inventories, including angiosperm and fern data, most of HERBase plots follow the terrain contour, while for fern data, most of HERBase plots follow the protocol by Tuomisto *et al.* (2003b) of installing plots along the topographic gradient. Despite their methodological differences, contour plots and plots along the topographic gradient can be combined if metadata on sub-plots is available to allow proper selection of comparable units (Moulatlet *et al.* 2017; Zuquim *et al.* 2019).